Suitability of Collagen Matrix as a Dural Graft in the Repair of Experimental Posterior Fossa Dura Mater Defects

DeneySEL Posterior Fossa Dura Mater DefekTleri Tamirinde Kollajen Matriksin Greft Olarak Uygunluğunun Incelenmesi

ABSTRACT
OBJECTIVE: To investigate the suitability of collagen matrix as a dural graft in the repair of experimental posterior fossa dura mater defects.

METHODS: Thirty New Zealand white rabbits were deployed in the study and randomly divided into two groups. In both groups, a 1 cm piece of dura mater was excised. The dura mater defect was covered by collagen matrix in the study group and the excised dura mater was sutured up to yield the pre-craniectomy condition in control group. After a follow-up period, the control dura mater and graft zone were excised together with the adjacent cerebellar cortex. Recovery and meningocerebellar adhesions were examined macroscopically and stained sections underwent light microscopy for detection of cellular inflammatory response, fibroblastic proliferation, foreign body reaction and capsule formation.

RESULTS: In the study group, a fibrous membrane indistinguishable from normal dura was observed macroscopically. There was significantly more fibroblastic proliferation in the study group compared to the control group in the first and second months. Foreign body reactions persisted in the control group whereas no reaction was detected in the study group in the third month and this difference was statistically significant. No capsule formation was observed in either group.

CONCLUSION: These findings suggest suitability of collagen matrix for duraplasty in the posterior fossa.

KEY WORDS: Collagen matrix, Duraplasty, Posterior fossa

ÖZ
AMAÇ: Bu çalışmada posterior fossada deneySEL olarak oluşturulan dura mater defekTlerinin tamirinde dural greft olarak kollajen matriksin uygunluğunu araştırdık.


SONUÇ: Bu bulgular kollajen matriksin posterior fossada duraplasti için uygun olabileceği kanaatine varmamızı sağladı.

ANAHTAR SÖZÇÜKLER: Arka çukur, Duraplasti, Kollajen matriks
INTRODUCTION

Repair of dural defects and dural closure without a leak is essential in reducing the risk of infection, brain herniation, cerebrospinal fluid (CSF) fistulae, meningocerebral adhesion and brain scarring (2).

Separation of the dura mater during prolonged posterior fossa surgery causes its retraction. In cases where cerebellar swelling is experienced or expected, primary closure is not possible due to the shrinkage of the edges of the dura mater or retraction of the dura making a dural graft necessary (4,15).

An experimental dura mater defect was created in the posterior fossa and suitability of collagen matrix as a graft was assessed.

MATERIAL and METHOD

The Ethical Committee of the Atatürk University Medical School approved the study protocol. Thirty New Zealand white rabbits weighing between 3.5 and 4.5 kg were used. The rabbits were divided into two groups as the study group and the control group. The animals were anesthetized with 5% isofluorane (Florane Liquid, Abbott Laboratories Ltd. Queensborough, Kent, UK) administered by a mask. Anesthesia was continued with 1-3% isofluorane inhalation. An occipitocervical midline incision was made and the bone was disclosed. A 1x1 cm piece of dura mater was excised with utmost care in order not to injure the pia-arachnoid membranes. The defect was covered with collagen matrix (Duragen, Integra, US) exceeding the edges and was soaked with 0.9% sodium chloride in the study group. In the control group, the excised dura mater piece was sutured in place with 6/0 silk suture material yielding the pre-craniectomy condition. Both groups were divided into three subgroups comprising of five rabbits with respect to the 1st, 2nd and 3rd follow-up months. The intervention sites were followed-up in all animals with regard to the development of CSF fistulae, hematoma, infection, and wound recovery.

All wounds recovered without any complications. Dura mater that was excised and re-sutured to its former position joined to the surrounding dura mater completely at the end of the first month in the control group; there was no change in these findings at the end of the second and third months. Collagen matrix began to be absorbed at the end of the first month in the study group and a fibrous membrane lighter and brighter than the intact dura mater developed in the implantation area. This newly formed membrane extended into the dura mater incessantly. Examination of the collagen matrix implantation area at the end of the second month in the study group revealed merging of the surrounding dura mater with the fibrous membrane tightly in a way that could not be distinguished from the dura mater morphologically. This newly formed membrane extended into the dura mater incessantly. Examination of the collagen matrix implantation area at the end of the second month in the study group revealed merging of the surrounding dura mater with the fibrous membrane tightly in a way that could not be distinguished from the dura mater morphologically. In the third month, collagen matrix had been absorbed completely and the newly formed fibrous membrane had blended in the surrounding dura mater, rendering it indistinguishable.
Although meningocerebellar adhesions were less common in the first month in the collagen matrix group, this finding was not significant (p = 0.490). None of these animals had adhesions in the second month and the difference between the two groups was not statistically significant (p = 0.292). Neither group had adhesions in the third month.

**Histopathological Findings**

In the first month, the inflammatory response consisted of mononuclear cells in the collagen matrix group. Although the inflammatory response was more marked in the collagen matrix group compared to the control animals, this difference was not statistically significant (p=0.766). Histopathological examination revealed profound invasion of the collagen matrix by fibroblasts, collagen beginning to accumulate in the collagen matrix and collagen matrix being absorbed (Figure 1). Fibroblastic proliferation was more marked in the collagen matrix group than in the control animals (p=0.030). In contrast, there was no significant difference between the two groups in terms of foreign body reactions (p = 0.197) (Figure 2).

The inflammatory response had decreased by the second month in both groups and mononuclear cells were predominant (Figure 3). Inflammatory responses were equivalent in both groups. The amount of collagen and absorption of the collagen matrix had increased in the collagen matrix group and fibroblastic proliferation had decreased in both groups. Fibroblastic proliferation was more marked in the collagen matrix group compared to the control group. Comparison of foreign body reactions yielded no statistical difference between the groups (p = 0.565) (Figure 2).

No inflammatory response was present in either group in the third month. Fibroblastic proliferation had decreased in both groups. Intense collagen accumulation was noted in the defect area in the collagen matrix group. Collagen matrix was

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![Figure 1](image1.png)

*Figure 1: Partial graft residue and granulation tissue in the implant area one month after the implantation of collagen matrix (H & E, x100).*

![Figure 2](image2.png)

*Figure 2: Means of inflammatory response, fibroblastic proliferation and foreign body reaction gradings in the control (A) and collagen matrix (B) groups.*

![Figure 3](image3.png)

*Figure 3: Mild mononuclear cell infiltration and prominent fibroblastic proliferation in the implant area 2 months after the collagen matrix implantation (H & E, x100).*
absorbed completely. No statistically significant difference was detected between the two groups with regard to fibroblastic proliferation ($p = 0.208$). While foreign body reaction persisted in the control group, there was no foreign body reaction in the collagen matrix group. The difference between the two groups was significant ($p = 0.038$) (Figure 2).

No capsule or neomembrane formation was present in either group throughout the follow-up periods.

**DISCUSSION**

Closure of a dural defect with fibrous tissue obtained from the operation area (pericranium or temporal fascia) is possible in many cases. Autografts such as pericranium and temporal fascia have many advantages as they are economical, nontoxic and easy to use. They possess excellent biological activities since they are non-immunogenic. Use of the patient’s own fascial lath is not a popular procedure due to the requirement for an additional incision, prolongation of the operation time, complications in the donor area, meningoencephal adhesions, and the reluctance of the patients to have a second scar (2,16,20). Although temporal fascia and pericranium are ideal graft materials, they may not be available or obtained in adequate amounts or they may have been disrupted due to trauma or during a surgical intervention. Homologous, heterologous or synthetic graft materials are used in such conditions.

Inert and non-resorbable materials used for closing dural defects have considerable disadvantages as they are foreign bodies, cause intense inflammatory reactions, induce unusual neovascularization with late hemorrhages and hematomas in the long term and cause neural compression (10). Collagen is a resorbable material that has been tested in various forms such as film, layer and sponge (3,12,15). The results of our study indicate that collagen matrix is absorbed completely in the implantation region, and that the fibrous membrane blends into the surrounding dura mater rendering it indistinguishable from the normal dura mater. Elimination of collagen matrix by complete absorption was confirmed microscopically.

The porous nature of the collagen matrix seems to promote fibroblastic proliferation and facilitate dural repair (15). In addition, collagen itself is known to have a chemotactic interaction with fibroblasts. This interaction may aid in the recolonization of the collagen implant (6,18). Dural repair with tight and usually synthetic materials is performed around the graft (8,18). Closure of the dural defect with these materials depends on the capsulation membrane formed by fibroblasts (21).

Since the collagen matrix is porous, dural repair initiates in and proceeds through the graft. Thus, the risk of the capsule enclosing the collagen matrix is low. The porosity of the graft seems to be an essential feature in dural repair (15). Porosity of the graft material is not only responsible for its feasibility but it also allows invasion of the graft by fibroblasts to repair the tissue (11). In our experimental study, the collagen matrix was invaded by fibroblasts to varying degrees due to its porous nature in all animals.

Since the collagen matrix is a graft material used to cover the dura mater defect, the operation period is short. Suturing the graft to the edges of the dura may cause a CSF leak from the needle holes. Collagen has the advantage of eliminating this complication.

In the disruption of the pia-arachnoid layer, meningoencephal adhesions form regardless of the dural repair technique (17). While care was taken not to injure the pia-arachnoid membrane, minimal damage to the arachnoid was present in the animals with adhesions. The possibility of operative injury increases in the presence of cortical adhesions. Thus, graft material to be used for dural repair should either not produce cortical adhesions or form minimal ones. The finding that the number of adhesions formed by the collagen matrix was similar to the ones in the control group that were closed by their own dura maters is significant.

The fact that previously used collagen layers and collagen films caused an inflammatory response, neomembrane formation and meningoencephal adhesion possibly due to the tanning procedure, has created a lack of confidence in their use (7,9,12,14).

We suggest that the foreign body reaction persisting through the third month in the control group is due to the silk suture material used. The nonexistence of capsule and neomembrane formation in the collagen matrix group may be attributed to the porous nature of the collagen matrix and the use of the collagen matrix fibers by fibroblasts as support in the repair process (14). However, repair with many semi-synthetic materials develops around the graft and the graft is
encapsulated (1,8,13,19). Capsule and neomembrane formation may thicken and in the long run, cause neural compression and hemorrhage due to its fragile capillaries (1,5,22).

The shaping of the collagen matrix in the posterior fossa was easily achieved without the use of a time-consuming suturing process in our experimental study. Collagen matrix was found to tolerate permanent cerebellar swelling without increasing the risk of CSF leakage in cases with no hydrocephalus and if the wound is closed carefully layer-by-layer (15).

Collagen matrix seemed to be absorbed completely and was used easily without any suturing. Foreign body reactions were minimal and disappeared in the long term. The intensity of the inflammatory response to collagen matrix was similar to that in the control group. Since there was no pia-arachnoid injury, adhesions did not form. Collagen matrix joined the dura mater and allowed its regeneration successfully. Since neomembrane or graft capsulation was not present owing to the porous nature of the matrix and the use of collagen fibers by fibroblast as a support, hemorrhage from neomembranous fragile capillaries is unlikely.

This study indicates that collagen matrix may be suitable for posterior fossa duraplasty. This technique may be particularly useful in patients in whom primary closure of the dura is not possible due to shrinkage, retraction and excision, or in conditions where the dura should be left open to allow the development of edema.

REFERENCES